

of intra-herd and inter-herd *S. aureus* strains. DNA-based methods are used to compare *S. aureus* isolates from staphylococci infections with human and animal origin and to describe epidemiologically (Kapur *et al.*, 1995; Zadoks *et al.*, 2000; Reinoso *et al.*, 2004). Enterobacterial repetitive intergenic consensus-polymerase chain reaction (ERIC) method has been determined to be reliable in identification of *Staphylococcus* and *Streptococcus* strains with human and animal origin (del Vecchio *et al.*, 1995; van Belkum *et al.*, 1996; Wieser and Busse, 2000; Reinoso *et al.*, 2007). ERIC-PCR has become a powerful tool for molecular genetic analysis of bacteria and bacterial taxonomy since it allows individual fingerprints of genus, species and strains and helps to determine the phylogenetic relationships (de Bruijn, 1992).

Studies about epidemiology, ecology, pathogenesis and strain variations of important *Staphylococcus* species significant udder pathogens in many countries and herds are limited. In this study, it is aimed to type advanced-level 98 *S. aureus* isolates isolated from the bovine mastitis by ERIC-PCR method.

MATERIALS AND METHODS

Bacterial strains

A total of 98 isolates from Konya Region were isolated from bovine subclinical mastitis milk samples. All isolates were identified as *S. aureus* by a standart microbiological procedure (Holt *et al.*, 1994) and confirmed by use of identification test with the VITEK 2 system (bioMérieux).

DNA isolation and PCR amplifications

Bacterial strains were incubated in Brain Heart Infusion Broth at 37 °C for 18 hours. Genomic DNA isolation was made modifying the method described by Ausubel *et al.* (1991).

DNA samples were amplified by PCR for the repetitive element sequence using the primer ERIC-PCR: ERIC1R: 5'-ATGTAAGCTCCTGGGGATTAC-3', ERIC2: 5'-AAGTAAGTGACTGGGGTGAGC-3' (Louws *et al.*, 1994; Versalovic *et al.*, 1991).

Each 25µl of the reaction contained 50ng of genomic DNA, 10XPCR reaction buffer (50mM KCl, 10mM Tris-HCl, pH=9, %0.1 TritonX-100), 3mM MgCl₂, 2.5mM dNTP, 75pmol of each primer and 2.5U of *Taq* DNA polymerase (Fermentas). PCR was performed in a Mastercycler Gradient thermal cycler (Eppendorf). The initial step of 95°C for 7 min was followed by 30 cycles of 94°C for 1 min, annealing 52°C for 1 min, 65°C for 8 min, and a final cycle at 65°C for 16 min. The amplification products were separated by gel electrophoresis in 2% agarose gels, stained with ethidium

bromide, photographed on a UV transilluminator. Gel image has been transferred to the computer with DNA imaging system (Vilber Lourmat).

Data analysis

Fingerprints were scored visually as present (1) or absent (0). Genetic similarity among *S. aureus* strains was estimated based on Nei homology using Bio1D++ computer programme. Cluster analysis was performed using the UPGMA.

RESULTS

Genetic relationship within all *S. aureus* isolates was analyzed by ERIC-PCR using the ERIC1R and ERIC2 primers. ERIC-PCR produced genomic profiles consisted of 8 to 14 bands, with a size of range of ~8000 bp and 250 bp.

According to the dendrogram obtained by the assessment of 28 bands in total (Fig.1), a differentiation was formed by analyzing the dendrogram with a similarity coefficient of 68% and considered to define six clusters namely I to VI. ERIC-PCR grouped 98 *S. aureus* isolates into these clusters I–VI showing 64 ERIC genotypes with discrimination indexes (*D*) of 0.96. While this result showed that genotypes were highly similar between *S. aureus* isolates, the discriminatory power by ERIC-PCR method was determined to be high. In the entire *S. aureus* isolates worked on, it was observed that two band patterns with sizes <6000 bp and 500 bp are common (monomorphic). In sizes 3000 bp, 1000 bp and 250 bp, common bands were observed in most isolates. It was found that the isolates in cluster VI exhibited their specific band profiles (2000 bp and 7500bp).

DISCUSSION

In identification of bacteria strains phenotypic characteristics such as biotyping, phage typing, serotyping and antibiotic resistance are used. However, more recently, molecular approaches are beneficial in genotypic distinction of bacteria strains. Restriction fragment length polymorphism (RFLP) (Owen, 1989), Pulsed field gel electrophoresis (PFGE) (Murray *et al.*, 1990; Gardella *et al.*, 2005), plasmid profiles (Litwin *et al.*, 1991; Arslan *et al.*, 2009a), repetitive extragenic palindromic-polymerase chain reaction (REP-PCR) (Versalovic *et al.*, 1994; Louws *et al.*, 1996; Arslan *et al.*, 2009b; Aruna *et al.*, 2009; Nordin *et al.*, 2010; Manga and Vyletřlová, 2012; Njage *et al.*, 2013; Kang and Dunne, 2003) and random amplified polymorphic DNA-Polymerase chain reaction (RAPD-PCR) (Arslan *et al.*, 2005; Gardella *et al.*, 2005) can be given as examples of some molecular genotyping methods.

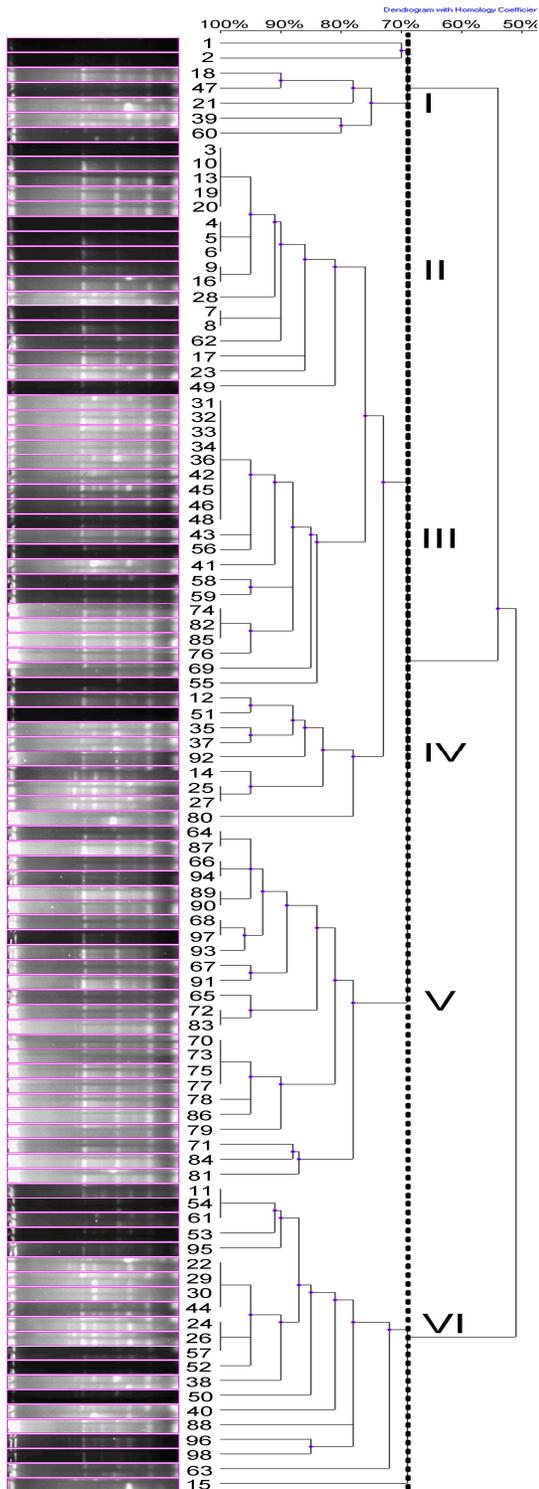


Fig. I. Genetic relationship among the *S. aureus* strains estimated by clustering analysis of DNA fingerprints generated by ERIC-PCR methods.

ERIC-PCR have been successfully used by many researchers in distinction of gram (-) (Versolavic *et al.*, 1991; Rodriguez-Barradas *et al.*, 1995; Kerouanton *et al.*, 1996; Appuhamy *et al.*, 1997; Diab and Al-Turk, 2011) and gram (+) (van Belkum *et al.*, 1993; Lipman *et al.*, 1996; Kang and Dunne, 2003) bacteria strains. Lipman *et al.* (1996), by genotyping of *S. aureus* strains from isolated bovine mammary glands, determined origin of outbreak of *S. aureus* in herd. It was emphasized that ERIC-PCR allow the clear differentiation of *Staphylococcus epidermidis* and *Staphylococcus hominis* strains which have a problem in biochemical tests by exhibiting species specific banding patterns (Wieser and Busse, 2000). By separating to 11 types with ERIC-PCR of 67 *S. aureus* strains, it was shown that ERIC-PCR technique provides a reliable tool for investigating epidemiology and tracking the spread of *S. aureus* strains in the hospital environment (Candan *et al.*, 2013). But in this study, ERIC-PCR showed better discrimination power with 64 genotypes of 98 *S. aureus* strains. The result of a previous study exhibiting 75 ERIC-types in 90 *S. aureus* was also in concordance with our data (Abdollahi *et al.*, 2014). As similar with our discrimination indexes (D) of 0.96, Ye *et al.* (2012) classified 35 *S. aureus* isolates into 28 ERIC types with discrimination indexes (D) of 0.984. It was shown that PFGE allowed better discrimination of *S. epidermidis* isolates than REP-PCR and RAPD methods (Begović *et al.*, 2013). Gardella *et al.* (2005) determined that ERIC method is as good as PFGE technique.

CONCLUSION

According to these studies, the ERIC-PCR method has been successfully applied for typing and distinguishing of *Staphylococcus* species and strains. In conclusion, it will be useful to create a fingerprint database through this rapid and reliable molecular method (ERIC-PCR) in finding the source of the epidemic causing the disease, determining the pathogen routinely and in vaccine development studies. In this way, rapid diagnosis will be possible and the treatment process will be shortened by providing useful data in developing effective protection strategies against the mastitis.

ACKNOWLEDGEMENTS

This research was carried out at the laboratory Molecular Biology, Selçuk University and was supported by the Scientific and Technological Research Council of

Turkey (TUBITAK, Project No:108T290). We would like to thank Prof. Dr. U. Sait UÇAN for his help in bacterial identification and for collection of the milk.

Statement of conflict of interest

Authors have declared no conflict of interest.

REFERENCES

- Aarestrup, F.M., Wegener, H.C. and Rosdahl, V.T., 1995. Evaluation of phenotypic and genotypic methods for epidemiological typing of *Staphylococcus aureus* isolates from bovine mastitis in Denmark. *Vet. Microbiol.*, **45**:139-150.
- Abdollahi, S., Ramazan-zadeh, R., Kalantar, E. and Zamani, S., 2014. Molecular epidemiology of *Staphylococcus aureus* with ERIC-PCR method. *Bull. Env. Pharmacol. Life. Sci.*, **3**: 158-165.
- Appuhamy, S., Parton, R., Coote, J.G. and Gibbs, H.A., 1997. Genomic fingerprinting of *Haemophilus somnus* by a combination of PCR methods. *J. clin. Microbiol.*, **35**: 288–291.
- Arslan, E., Açıık, L. and Uçan, U.S., 2005. Determination by RAPD-PCR of Relationship Degrees of *S. aureus* and *S. intermedius* Strains Isolated from Bovine Mastitis. *S.Ü. J. Vet. Sci.*, **21**: 65-69.
- Arslan, E., Çelebi, A., Açıık, L. and Uçan, U.S., 2009a. Characterization of coagulase positive *Staphylococcus* species isolated from bovine mastitis by protein and plasmid patterns. *Turk. J. Vet. Anim. Sci.*, **33**: 493-500.
- Arslan, E., Ertuğrul, K. and Albayrak, A.İ., 2009b. Typing of *Staphylococcus aureus* and *Staphylococcus intermedius* strains with repetitive sequence-based PCR (REP-PCR) method. *S.Ü. Fen. Fak. Fen. Derg.*, **34**: 85-92.
- Aruna, V., Dhanalakshmi, K., Sarma, B.J.R. and Anjaneyulu, Y., 2009. Characterization of *Staphylococcus aureus* isolates by REP and BOX PCR methods. *Indian. Vet. J.*, **86**: 1-3.
- Ausubel, F.M., Kingston, R.E., Brent, R., Moore, D.D., Seidman, J.G., Smith, J.A. and Struhl, K., 1991. *Current protocols in molecular biology*. Greene Publishing Associates & Wiley Interscience, New York.
- Begović, J., Jovčić, B., Papić-Obradović, M., Veljović, K., Lukić, J., Kojić, M. and Topisirović, L., 2013. Genotyping diversity and virulent factor of *Staphylococcus epidermidis* isolated from human breast milk. *Microbiol. Res.*, **168**: 77-83.
- Candan, E.D., Idil, N. and Seyis-Bilkay, I., 2013. Usefulness of REP and ERIC-PCR combination for tracking the spread of *Staphylococcus aureus* strains. *Minerva. Biotechnol.*, **25**: 245-50.
- de Bruijn, F.J., 1992. Use of repetitive (repetitive extragenic palindromic and enterobacterial repetitive intergenic consensus) sequences and the polymerase chain reaction to fingerprint the genomes of *Rhizobium meliloti* isolates and other soil bacteria. *Appl. environ. Microbiol.*, **58**: 2180-2187.
- Del Vecchio, V.G., Petroziello, J.M., Gress, M.J., McClesky, F.K., Melcher, G.P., Crouch, H. K. and Lupski, J. R., 1995. Molecular genotyping of methicillin-resistant *Staphylococcus aureus* via flu-orphore enhanced repetitive sequence VCR. *J. clin. Microbiol.*, **33**:2141-2144.
- Diab, A.M. and Al-Turk, I.M., 2011. ERIC and RAPD PCR-based DNA fingerprinting techniques application for microbial source tracking (MST) at Al-Madinah Al-Munwarrah, KSA. *J. Taibah Univ. Sci.*, **5**: 31-38.
- Erganiş, O., Kuyucuoğlu, Y. and Ok, Ü., 1995. İnek ve koyun mastitislerine sebep olan koagulaz negatif ve pozitif Stafilokokların biyotiplendirilmesi. *Veterinarium*, **6**: 23-27.
- Gardella, N., Picasso, R., Predari, S.C., Lasala, M., Foccoli, M., Benchetrit, G., Famiglietti, A., Catalano, M., Mollerach, M. and Gutking, G., 2005. Methicillin-resistant *Staphylococcus aureus* strains in Buenos Aires Teaching Hospitals: replacement of the multidrug resistant South American clone by another susceptible to rifampin, minocycline and trimethoprim-sulfamethoxazole. *Rev. Argent. Microbiol.*, **37**: 156–160.
- Gudding, R., McDonald, J.S. and Cheville, N.F., 1984. Pathogenesis of *Staphylococcus aureus* bacteriological, histologic and ultrastructural pathologic finding. *Am. J. Vet. Res.*, **45**: 2525-2531.
- Han, H.R., Pak, S-II., Kang, S.W., Jong, W.S. and Youn, C.J., 2000. Capsular polysaccharide typing of domestic mastitis causing *Staphylococcus aureus* strains and its potential exploration of bovine mastitis vaccine development, I. Capsular polysaccharide typing, isolation and purification of the strains. *J. Vet. Sci.*, **1**: 53-60.
- Holt, J.G., Krieg, N.R., Sneath, P.H.A., Staley, J.T. and Williams, S.T., 1994. In: *Bergey's manual of determinative bacteriology*. 9th edn. (William R Hensyl), Williams & Wilkins, Baltimore, pp. 518.
- Ibarra-Velázquez, L.M., Torres-Vitela, M.R., Andrade-González, I., López-Muraira, I.G., Valdés-Rodríguez, S. E. and Gómez-Leyva, J.F., 2011 Genetic variation and antibiotic susceptibility among *Staphylococcus aureus* isolates from dairy products and food handlers. *Afr. J. microbiol. Res.*, **5**: 4901-4908.
- Kang, H.P. and Dunne, W.M., 2003. Stability of rep-PCR patterns with respect to culture age and subculture frequency. *J. clin. Microbiol.*, **41**: 2694-2696.
- Kapur, V., Sischo, W., Greer, R., Whittam, T. and Musser, J., 1995. Molecular population genetic analysis of *Staphylococcus aureus* recovered from cows. *J. clin. Microbiol.*, **33**: 376-380.
- Kerouanton, A., Brisabois, A., Grout, J. and Picard, B., 1996. Molecular epidemiological tools for *Salmonella dublin* typing. *FEMS. Immunol. med. Microbiol.*, **14**: 25–29.

- Lipman, L.J.A., de Nijs, A., Lam, T.J.G.M., Rost, J.A., Van, D.I.J.K.L., Schukken, Y.H. and Gaastra, W., 1996. Genotyping by PCR of *Staphylococcus aureus* strains isolated from mammary glands of cows. *Vet. Microbiol.*, **48**: 51-55.
- Litwin, C.M., Storm, A.L., Chipowsky, S. and Ryan, K.J., 1991. Molecular epidemiology of Shigella infections: plasmid profiles, serotype correlation, and restriction endonuclease analysis. *J. clin. Microbiol.*, **29**: 104-108.
- Louws, F.J., Fulbright, D.W., Stephens, C.T. and de Bruijn, F.J., 1994. Specific genomic fingerprints of phytopathogenic *Xanthomonas* and *Pseudomonas pathovars* and strains, generated with repetitive sequences and PCR. *Appl. environ. Microbiol.*, **60**: 2286-2295.
- Louws, F.J., Schneider, M. and de Bruijn, F.J., 1996. In: *Nucleic acid amplification methods for the analysis of environmental samples* (ed. G. Toranzos). Technomic Publishing Co, pp. 63-94.
- Manga, I. and Vyletřlová, M., 2012. Rep-PCR-based typing as a tool for tracking of MRSA infection origin. *Acta Univ. Agric. Silvicult. Mendel. Brun.*, **40**: 251-256.
- Murray, B.E., Singh, K.V., Heath, J.D., Sharma, B.R. and Weinstock, G.M., 1990. Comparison of genomic DNAs of different enterococcal isolates using restriction endonucleases with infrequent recognition sites. *J. clin. Microbiol.*, **28**: 2059-2063.
- Njage, P.M.K., Dolci, S., Jans, C., Wangoh, J., Lacroix, C. and Meile, L., 2013. Biodiversity and enterotoxigenic potential of staphylococci isolated from raw and spontaneously fermented camel milk. *Br. microbiol. Res. J.*, **3**: 128-138.
- Nordin, S., Wahab, Z., Hussin, S. and Rahman, M., 2010. Molecular characterization of nosocomial methicillin resistant *staphylococcus aureus* by Rep-PCR. *Internet. J. Infect. Dis.*, **9**: 2.
- Olive, D.M. and Bean, P., 1999. Principles and applications of methods for DNA-based typing of microbial organisms. *J. clin. Microbiol.*, **37**: 1661-1669.
- Qayyum, A., Jawaria, A. K., Riaz, H., Muhammad, A., Nisar, A., Ahrar, K. and Muhammad, S. K., 2016. Prevalence and Association of Possible Risk Factors with Sub-Clinical Mastitis in Cholistani Cattle. *Pakistan J. Zool.*, **48**: 519-525.
- Owen, R.J., 1989. Chromosomal DNA fingerprinting—a new method of species and strain identification applicable to microbial pathogens. *J. med. Microbiol.*, **30**: 9-99.
- Reinoso, E., Bettera, S., Frigerio, C., Direnzo, M., Calzolari, A. and Bogni, C., 2004. RAPD-PCR analysis of *Staphylococcus aureus* strains isolated from bovine and human hosts. *Microbiol. Res.*, **159**: 245-255.
- Reinoso, E., Bettera, S., Odierno, L., and Bogni, C., 2007. rep-PCR of *Staphylococcus aureus* strains isolated from bovine mastitis in Argentina. *Braz. J. Vet. Res. Anim. Sci.*, **44**: 115-121.
- Rodriguez-Barradas, M.C., Hamill, R.J., Houston, E.D., Georghiou, P.R., Clarridge, J.E., Regnery, R.L. and Koheler, J.E., 1995. Genomic fingerprinting of *Bartonella* species by repetitive element PCR for distinguishing species and isolates. *J. clin. Microbiol.*, **33**: 1089-1093.
- Singh, M.P. and Singh, C.M., 1968. Fungi isolated from clinical cases of bovine mastitis in India. *Indian J. anim. Hlth.*, **7**: 259-263.
- Struelens, M.J., 1996. Consensus guidelines for appropriate use and evaluation of microbial epidemiologic typing systems. *Clin. Microbiol. Infect.*, **2**: 2-11.
- Tenover, F.C., Arbeit, R., Archer, G., Biddle, J., Byrne, S., Goering, R., Hancock, G., Herbert, G.A., Hill, B., Hollis, R., Jarvis, W.R., Kreiswirth, B., Eisner, W., Maslow, J., McDougal, L.K., Miller, J.M., Mulligan, M. and Pfaller, M.A., 1994. Comparison of traditional and molecular methods of typing isolates of *Staphylococcus aureus*. *J. clin. Microbiol.*, **32**: 407-415.
- Tenover, F.C., Arbeit, R.D., Goering, R.V., Mickelsen, P.A., Murray, B.E., Persing, D.H. and Swaminathan, B., 1995. Interpreting chromosomal DNA restriction patterns produced by pulsed-field gel electrophoresis: criteria for bacterial strain typing. *J. clin. Microbiol.*, **33**: 2233-2239.
- Van Belkum, A., Bax, R., Peerbooms, P., Goessens, W.H.F., Van Leeuwen, N. and Quint, W.G.V., 1993. Comparison of phage typing and DNA fingerprinting by polymerase chain reaction for discrimination of methicillin-resistant *Staphylococcus aureus* strains. *J. clin. Microbiol.* **31**: 798-803.
- Van Belkum, A., Sluijter, M., de Groot, R., Verbrugh, H. and Hermans, P.W., 1996. Novel BOX repeat PCR assay for high-resolution typing of *Streptococcus pneumoniae* strains. *J. clin. Microbiol.*, **34**: 1176-1179.
- Van Belkum, A., Tassios, P.T., Dijkshoorn, L., Haeggman, S., Cookson, B., Fry, N.K., Fussing, V., Green, J., Feil, E., Gerner-Smidt, P., Brisse, S. and Struelens, M., 2007. Guidelines for the validation and application of typing methods for use in bacterial epidemiology. *Clin. Microbiol. Infect.*, **13**: 1-46.
- Versalovic, J., Schneider, M., de Bruijn, F.J. and Lupski, J.R., 1994. Genomic fingerprinting of bacteria using repetitive sequence based PCR (rep-PCR). *Meth. Mol. Cell. Biol.*, **5**: 25-40.
- Versalovic, J., Koeuth, T. and Lupski, J.R., 1991. Distribution of repetitive DNA sequences in eubacteria and application to fingerprinting of bacterial genomes. *Nucl. Acid. Res.*, **19**: 6823-6831.
- Wieser, M. and Busse, H.J., 2000. Rapid identification of *Staphylococcus epidermidis*. *Int. J. Syst. Evol. Microbiol.*, **50**: 1087-1093.
- Ye, Y., Jiang, Q., Wu, Q., Zhang, J., Lu, J. and Lin, L., 2012. The characterization and comparison of *Staphylococcus aureus* by antibiotic susceptibility testing, enterobacterial repetitive intergenic consensus-polymerase chain reaction, and random amplified polymorphic DNA-polymerase chain reaction. *Foodborn. Pathogen. Dis.*, **9**: 168-171.

- Zadoks, R., Van Leeuwen, W., Barkema, H., Sampimon, O., Verbrugh, H., Schukken, Y. and Van Belkum, A., 2000. Application of pulsed-field gel electrophoresis and binary typing as tools in veterinary clinical microbiology and molecular epidemiologic analysis of bovine and human *Staphylococcus aureus* isolates. *J. clin. Microbiol.*, **38**: 1931-1939.
- Zadoks, R.N., Middleton, J.R., McDougall, S., Katholm, J. and Schukken, Y.H., 2011. Molecular epidemiology of mastitis pathogens of dairy cattle and comparative relevance to humans. *J. Mamm. Gl. Biol. Neoplasia*, **16**: 357-372.